

slower rates in about 8 days after PI, secondary branch primordia started developing on primary branch at the tip. The sequence of development of primary and secondary branch primordia gradually progressed downwards. Thus, the development of spikelets also took place in basipetal order. By about 12 days, glumes, lemma and palea were developed at the tip but these were less developed at the middle of the panicle. At the base, only glumes were to be developing. After about 16 days, all the floral parts, except anthers and stigmas, were fully developed at the tip but less developed at the middle. Within 20 days, the ovary was fully developed, while the stigmas just started developing. At the middle and the base of the panicle, only anthers were developing; stigmas had not yet initiated. By 22 days, all the floral parts were fully developed at the tip, middle and the base of the panicle. Maiti (1977) reported that the development of panicle components in the hybrids superceded their parents. The hybrids and their parents have therefore shown distinct patterns in the length of vegetative period and rate of primordia differentiation. The high yielding capability of the hybrids was reflected in their superiority at an early stage of panicle development. Similar studies in other hybrids may validate this conclusion. Rao and Venkateswarlu (1976) and Gibson and Schertz (1977) have also shown the superiority of hybrids over their parents in terms of panicle development.

Seasonal difference:

The development time of different phenological stages of panicle is short in the rainy season, longer in early post-rainy season and still longer in late post-rainy season in Hybrid 22E, which was very early maturing, showing considerable deviation from its parents in all the seasons. The developmental timetables of CSH1 did not deviate from 22E during the rainy season, but 22E showed much deviation from CSH1 in late post-rainy season. However, development of hybrid 22E was early in all seasons when compared to CSH1 (Fig. 5.12-5.13). The hybrids showed considerable decline in growth components in late post-rainy season compared to that in the rainy season (in 1976, India). The dry weight of panicle of CSH1 and 22E showed steep rise from 36-48 days after panicle initiation beyond which there was no significant increase in growth in rainy season. In late post-rainy season, growth of the cultivars declined considerably but increased again 52 days after PI.

Development and maturity of grain:

The grain is the ripened ovary with attached glumes. During development and maturation, grains pass through several distinct phases. The process starts with the formation of waxy fluid in the grain which is gradually condensed to the milky white stage. This in turn is converted to soft, and finally to hard endosperm stage. The grain growth terminates with the formation of black layer at the hilar region. The initiation of black layer shows a semi-lunar brownish ring which gradually encircles the hilum and gradually converts it back to a black layer. Phloem parenchyma are blocked with mucilage and pectic compounds at maturity and form a black layer (Quinby, 1972a).

The structure and ontogeny of the black layer has been studied by Giles *et al.* (1976). The early appearance of phenolic compounds in the cells of the phloem parenchyma is accompanied with the formation of the dark patch adjacent to the transfer cells. Mucilage, possibly arising from the breakdown of slime strands, represents slime plugs in the sieve tubes. The appearance of mucilage coincided with the formation of pectic compounds and callose indicate the senescence of the phloem tissue and the cessation of assimilate translocation. The xylem gets separated from the phloem just below the lodicules, and the phloem forms a band which continues into the pericarp on the abgerminal side. The centio-chalazal pad lying between the band of phloem parenchyma and the transfer cells is made up of thin walled isodiametric cells and is neither crushed or compressed. The black layer appears as a brown band of tissue near the basal abgerminal side of pericarp in the area of transfer cells.

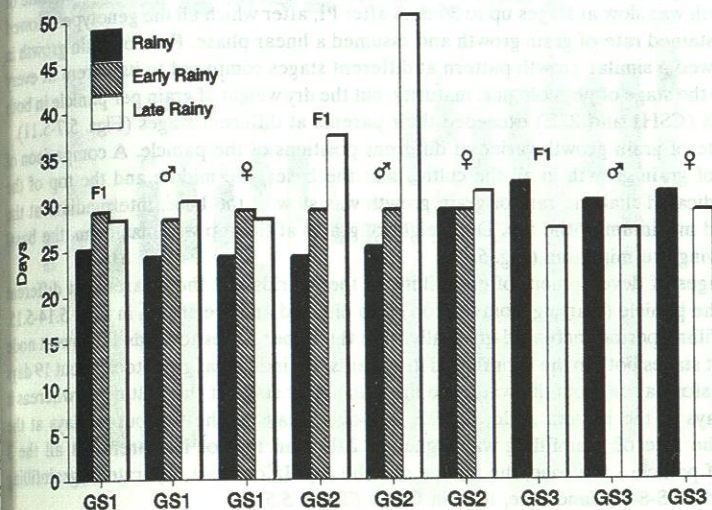


Figure 5.12 Growth stages of hybrid CSH1 and its parents in different growing seasons (Maiti, 1977).

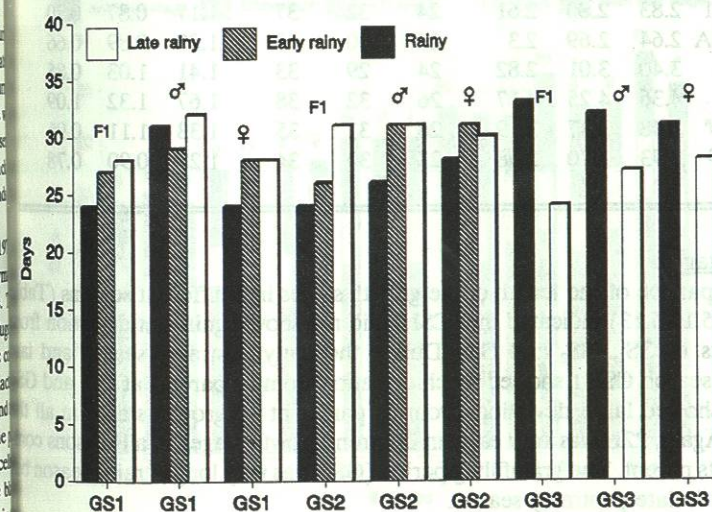


Figure 5.113 Growth stages of hybrid 22E and its parents in different growing seasons (Maiti, 1977).

A comparative study in rainy season on the grain dry matter (on the basis of 100 g weight) at different stages of both CSH1, 22E and their parents indicated that the rate of grain growth was slow at stages up to 36 days after PI, after which all the genotypes showed a more sustained rate of grain growth and assumed a linear phase. Rate of grain growth in CSH1 showed a similar growth pattern at different stages compared to its parent at the top stage upto the stage of physiological maturity, but the dry weight of grain per panicle in the hybrids (CSH1 and 22E) exceeded their parents at different stages (Figs. 5.7-5.11).

The rate of grain growth varied at different positions of the panicle. A comparison of the rates of grain growth in all the cultivars at the bases, the middle and the top of panicle indicated that the rate of grain growth was slow at the base, intermediate at middle and maximum at the top. Dry weight of grains at the top was maximum, the middle showing the minimum (Fig. 5.11).

The stages of development of grainfilling in the hybrids and their parents at different nodes of the panicle (starting from the top) also differed and are shown in Figs. 5.14-5.15. The grainfilling period increased gradually from the upper nodes towards the lowest nodes at different stages both in the hybrids and its parents. An individual grain took about 19 days for its transformation from the watery to the black layer stage at the first node, whereas it took 26 days at the bottom node. In 22E, it took 18 days at the top but 26 days at the bottom. The rate of grainfilling was higher in 22E than that of its parents at all the portions of panicle - the base, the middle and the top. In contrast, the rate of grainfilling was higher in IS-84, a land race, than in CSH1 (Table 5.5).

Table 5.5 Grain filling period and grain filling rate of CSH-1, 22E and their parents at different locations of panicle (top, middle, base) during rainy season 1975 (Maiti, 1977).

| Genotype | 100 seed weight(g) | | | Grain filling period (days) | | | Grain filling rate | | |
|----------|--------------------|--------|------|-----------------------------|--------|------|--------------------|--------|------|
| | Top | Middle | Base | Top | Middle | Base | Top | Middle | Base |
| CSH-1 | 2.83 | 2.80 | 2.61 | 24 | 32 | 37 | 1.17 | 0.87 | 0.70 |
| CK60A | 2.64 | 2.69 | 2.3 | 22 | 30 | 35 | 1.20 | 0.89 | 0.66 |
| IS-84 | 3.40 | 3.01 | 2.82 | 24 | 29 | 33 | 1.41 | 1.03 | 0.85 |
| 22E | 4.36 | 4.25 | 4.17 | 26 | 32 | 38 | 1.67 | 1.32 | 1.09 |
| 22E ♂ | 3.48 | 3.47 | 3.19 | 26 | 31 | 35 | 1.33 | 1.11 | 0.91 |
| 22E ♀ | 2.93 | 2.70 | 2.68 | 23 | 30 | 34 | 1.27 | 0.90 | 0.78 |

Growth stages:

A comparison of the length of the growth stages in 4 different seasons (Table 5.6; Figs. 5.12-5.13) indicated that CSH1 did not show significant deviation from its parents in GS₁, GS₂ and GS₃. During the early post-rainy season and late post-rainy season CSH1 showed much deviation from its parents at GS₂ and GS₃ but 22E showed large deviations from its parent at all growth stages in all seasons. Again, 22E was very early in different growth stages in all seasons compared to its parent. The grainfilling period (GS₃) was very long in rainy season and very short in late post-rainy season.

Effect of weather on growth stage:

Table 5.7 shows the meteorological conditions under 3 stages for the 3

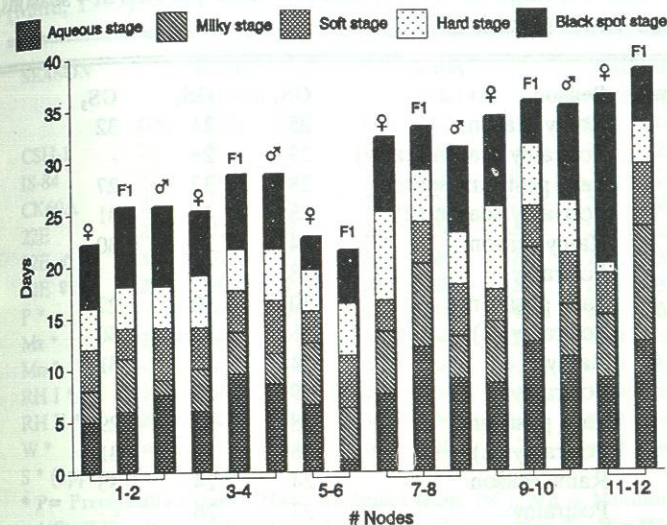


Figure 5.14 Grain filling stage of hybrid CSH1 and its parents in different nodes of the panicle during the rainy season 1975 (Maiti, 1977).

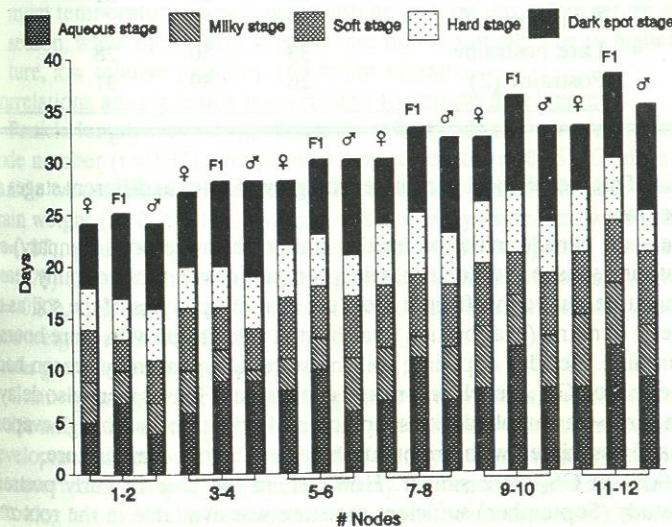


Figure 5.15 Grain filling stage of hybrid 22E and its parents in different nodes of the panicle during the rainy season 1975 (Maiti, 1977).

Table 5.6 Growth stages of CSH1, 22E and their parents in 4 seasons at Patancheru, India (Maiti, 1977).

| Genotype | Season | GS ₁ | GS ₂ | GS ₃ |
|----------|--------------------------|-----------------|-----------------|-----------------|
| CSH1 | Rainy season | 25 | 24 | 32 |
| | Postrainy season (early) | 29 | 28 | - |
| | Late postrainy season | 28 | 37 | 27 |
| | Postrainy season (2) | 28 | 41 | 31 |
| IS84 ♂ | Rainy season | 24 | 25 | 30 |
| | Postrainy | 28 | 29 | - |
| | Late postrainy | 30 | 50 | 27 |
| | Postrainy (2) | 35 | 38 | 30 |
| CK60A ♀ | Rainy | 24 | 28 | 31 |
| | Postrainy | 29 | 28 | - |
| | Late postrainy | 28 | 31 | 29 |
| | Postrainy (2) | 26 | 42 | 31 |
| 22E | Rainy season | 24 | 24 | 33 |
| | Postrainy | 27 | 26 | - |
| | Late postrainy | 28 | 31 | - |
| | Postrainy (2) | 24 | 35 | 33 |
| 22E ♂ | Rainy season | 31 | 26 | 32 |
| | Postrainy | 29 | 31 | - |
| | Late postrainy | 32 | 31 | 27 |
| | Postrainy (2) | 35 | 35 | 30 |
| 22E ♀ | Rainy | 24 | 28 | 31 |
| | Postrainy | 28 | 31 | - |
| | Late postrainy | 33 | 30 | 28 |
| | Postrainy (2) | 26 | 39 | 31 |

The effect of meteorological parameters on growth rates at different stages is discussed below:

GS₁: The delay in panicle initiation for the late postrainy seasons (January) and early postrainy season (September) trials compared with those of rainy season (June) might be due to insufficient moisture in the top layers of the soil as atmospheric demand (evaporation) was high in association with more hours of bright sunshine. Besides depleting soil moisture, the post rainy season had a delaying effect on GS₁. Development and expansion of leaves were also delayed, though the crop was supplied with supplemental irrigation due to high evaporative demand associated with bright sunshine and higher temperature.

GS₂: Conditions in GS₂ were similar. However, in the case of early postrainy season's study (September) sufficient moisture was available in the root zone and this ensured that the growth rate was not affected.

GS₃: Fewer hours of bright sunshine and associated low day temperatures were the main causes for the delay in growth rate in the case of rainy season trials

Table 5.7 Weather and growth stages of sorghum in different seasons (Maiti, 1977).

| SEASON | Summer 19.1.1976 | | | Rainy 14.6.1975 | | | Post rainy 11.9.1976 | |
|---------------|---------------------|-----------------|-----------------|--------------------|-----------------|-----------------|-------------------------|-----------------|
| | GS ₁ | GS ₂ | GS ₃ | GS ₁ | GS ₂ | GS ₃ | GS ₁ | GS ₂ |
| CSH-1 | 29 | 37 | 27 | 25 | 24 | 32 | 29 | 28 |
| IS-84 | 31 | 50 | 27 | 24 | 26 | 29 | 28 | 26 |
| CK60A | 29 | 31 | 29 | 24 | 27 | 32 | 29 | 28 |
| 22E | 29 | 31 | 24 | 24 | 24 | 34 | 27 | 26 |
| 22E ♂ | 33 | 32 | 29 | 31 | 26 | 32 | 26 | 31 |
| 22E ♀ | 29 | 30 | 26 | 24 | 28 | 32 | 28 | 31 |
| P * | 0 | 0.5 | 88 | 142 | 140 | 149 | 21 | 0.6 |
| Mx * | 26-30 | >30 | >35 | >30 | =30 | 25.3 | >30 | >30 |
| Mn * | <15 | <20 | =20 | >20 | >20 | >20 | >20 | 16 |
| RH I * | >80 | >65 | =50 | >75 | >85 | >90 | >80 | >75 |
| RH II * | 20.4 | <20 | 17 | <55 | >70 | >70 | >40 | =20 |
| W * | <10 | <10 | <10 | >20 | 10.2 | 5.2 | <10 | <10 |
| S * (approx.) | =9 | | | =5 | | | =8 | |

* P = Precipitation (mm); Mx = Maximum temp. (°C); Mn = Minimum temp. (°C); R.H. = Relative Humidity (1 = morning, 11 = evening; %), W = Wind speed (Km/h), S = Sunshine (hr/day).

as the moisture was unlimited in this phase. The duration of the grainfilling period appeared to be lengthened by the prevailing lower minimum and maximum temperatures, low bright sunshine and low day temperature in the rainy season, while the reverse was the case during summer due to higher temperature, low relative humidity and bright sunshine.

Correlations among various traits related to panicle development:

Panicle length showed significant ($P < 0.05$) positive correlation with panicle node number ($r=0.95$), and primary branch length ($r=0.85$). Grain number per panicle was significantly related to the secondary branch number ($r=0.94$), and grain weight ($r=0.79$). Head weight was positively associated with grain number ($r=0.87$), grain weight ($r=0.99$), husk weight ($r=0.83$), and 100 seed weight ($r=0.78$). Days to anthesis showed negative association with primary branch length ($r=-0.87$). GS₃ days were found to show positive association with seed size (100 seed weight; $r=0.85$; Maiti, 1977).

Grain growth pattern (general)

Grain ripening is characterised by grain growth which is associated with increase in size and weight, change in grain color and leaf senescence. The process of grain development starts with the formation of watery fluid in the grain which is gradually converted to milky white, soft and finally hard endosperm stages. Initial growth following fertilization is free nuclear division in the endosperm. Following cell wall formation, the endosperm increases in size (unpublished; see Chapter 2 for details).

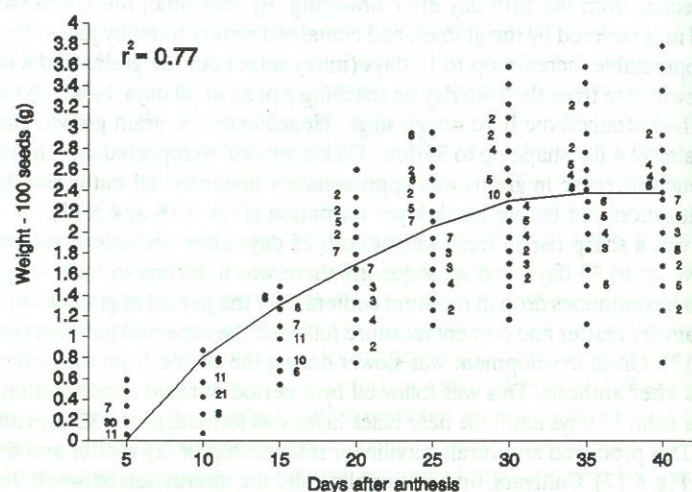


Figure 5.17 Accumulation of dry matter during the grain filling stage of (rainy season; Maiti *et al.*, 1979). ($n = 50$)

a set of 40 genotypes belonging to different taxonomic groups were chosen (Table 5.8). In order to study their behaviour in different seasons they were grown in early presummer (January) and postrainy season (September), 1976. The development of panicles from the date of initiation to physiological maturity was carefully observed, and the relationship between developmental stages and the yield components were also investigated. The mean number of days required for each developmental stage and yield components are given in Table 5.9.

Variability in developmental stages:

There was a wide range of variation in the vegetative and reproductive stages among various cultivars. For example, in the January experiment GS_1 ranged from 28 to 39 days, GS_2 from 24 to 64 days and GS_3 from 24 to 29 days. The number of leaves at panicle initiation ranged from 8 to 12, about 6-9 leaves were expanded at panicle initiation. Data on yield components also showed that the cultivars were widely variable. Panicle length ranged from 9.5 to 36.8 cm; node number from 12.5; number of primary branches from 95 to 71; number of secondary branches from 82 to 409; grains per panicle from 524 to 2307; grain weight per panicle 0.1 to 35 gm; and 100 seeds weight 0.55 to 4 gm. Out of 40 genotypes, only 20 flowered.

A close look at the developmental stages in both the seasons (Table 5.9) indicates that except for the three-leaf stage, all the developmental stages in the vegetative phase were delayed to a greater extent in the January planting than in the postrainy season crop. Stem elongation too was delayed in this season and

Table 5.8 Forty genotypes belonging to different taxonomic groups used in the development study.

| Taxonomic group | IS | Origin | Taxonomic group | IS | Origin |
|----------------------|-------|--------------------|--------------------|-------|------------|
| Bicolor | 714 | USA | Durra-Feterita | 7089 | Equatorial |
| Bicolor-Kafir | 658 | USA | Durra-Feterita | 7090 | Equatorial |
| Caffrorum | 183 | USA | Durra-Kafir | 3921 | USA |
| Caffrorum-Roxburghii | 2210 | USA | Durra-Kaura | 11025 | Ethiopia |
| Caudatum | 9743 | Sudan | Durra-Roxburghii | 4310 | India |
| Caudatum-Kafir | 127 | USA | Grass-grains | 1059 | India |
| Caudatum-Kaura | 7755 | Nigeria | Guinese | 3819 | Africa |
| Caudatum-Durra | 11574 | Ethiopia | Margaritifera | 8064 | Japan |
| Caudatum-Guinese | 3460 | Sudan | Milo-Kaura | 693 | USA |
| Caudatum-Nigricans | 8951 | Kenya | Nervosum | 11085 | Ethiopia |
| Cernum | 1054 | India | Nervosum | 11302 | Ethiopia |
| Conspicuum | 3818 | Africa | Nervosum Broomcorn | 11302 | USA |
| Conspicuum | 7999 | Nigeria | Nervosum-Kaoliang | 301 | USA |
| Conspicuum | 7630 | Nigeria | Roxburghii | 7276 | Nigeria |
| Dochna-Collier | 3648 | USA | | 7818 | Nigeria |
| Dochna-Amber | 601 | USA | | 6260 | India |
| Dochna-Honey | 633 | USA | Roxburghii-Shallu | 474 | USA |
| Dochna-Leoti | 640 | USA | Subglabrescens | 11150 | Ethiopia |
| Dochna-Nigricans | 2445 | USA | Zera-Zera | 3541 | Sudan |
| Durra | 4850 | India | | | |
| Shalepense | | Bangkok/Tetraploid | Bangkok | | |

Table 5.9 Mean estimates of number of days for different developmental stages in 2 seasons (40 genotypes).

| Stages | Postrainy season | | Late postrainy season | |
|------------------------|------------------|-------|-----------------------|-------|
| | Means | SD | Means | SD |
| 1. 3-Leaf stage | 4.65 | 0.48 | 4.22 | 1.25 |
| 2. 5-Leaf | 18.86 | 1.72 | 14.41 | 1.39 |
| 3. Panicle initiation | 30.68 | 9.76 | 30.70 | 2.56 |
| 4. Flag leaf | 50.15 | 9.86 | 58.11 | 10.95 |
| 5. Boot stage | 50.61 | 10.86 | 64.85 | 11.83 |
| 6. Half bloom | 66.76 | 10.21 | 72.22 | 11.25 |
| 7. Soft dough | 86.54 | 6.75 | 83.44 | 10.59 |
| 8. Hard dough | 93.59 | 8.72 | 89.11 | 10.90 |
| 9. Physiolog. maturity | 100.88 | 7.17 | 94.96 | 10.90 |
| GS_2 | 36.08 | 4.92 | 41.52 | 11.02 |
| GS_3 | 34.04 | 6.46 | 22.74 | 2.64 |

panicle initiation did not show any difference. Stem elongation took longer in January planting but the stages during grainfilling (GS₃, soft dough, hard dough and physiological maturity) were quite early. Seed size was reduced, which may lead to quick filling of the grain. Grainfilling also decreased to half during the postrainy season. The components of yield were lower in this season and affected panicle productivity (Table 5.10). This clearly reflected the effect of seasonal climate on crop phenology.

Table 5.10 Mean estimates of different yield components during the rainy season and the late postrainy season, 1976 (40 genotypes).

| Component | Postrainy SEASON | | Late postrainy | |
|---|------------------|-------|----------------|-------|
| | Means | SD | Means | SD |
| 1. Plant height (cm) | 174.88 | 6.89 | - | - |
| 2. Canopy height (cm) | 134.26 | 53.83 | - | - |
| 3. Stem elongation (days) | 26.93 | 4.63 | 34.15 | 11.44 |
| 4. Leaf number | - | - | 15.85 | 2.51 |
| 5. Flag leaf area (cm ²) | 95.04 | 7.59 | - | - |
| 6. Panicle length (cm) | 24.59 | 13.46 | 29.41 | 32.23 |
| 7. Panicle dry wt. (g) | 32.72 | 14.79 | - | - |
| 8. # secondary branches | - | - | 256 | 85 |
| 9. Grain number/panicle | - | - | 1100 | 476 |
| 10. Seed wt./panicle (g) | 25 | 16.46 | 14.80 | 9.68 |
| 11. 100 seed weight (g) | 2.67 | 1.17 | 1.96 | 0.79 |
| 12. Rate of grain filling per 100 seeds | 0.22 | 1.27 | 0.09 | 0.04 |

The effect of weather on growth stages:

When comparing the differences in the GS₁, GS₂ and GS₃ periods in different taxonomic groups between postrainy and late postrainy seasons, the following conclusions can be drawn (Table 5.11): 1- on the basis of lifespan, the cultivars may be tentatively divided into 4 groups, A, B, C and D; 2- in all cases, GS₁ was longer in the late postrainy season; 3- in group C, GS₂ was longer during the late postrainy season; 4- in group D, GS₂ was longer during the postrainy season; 5- duration of GS₂ increased with high temperature and low relative humidity, bright sunshine hours and clear skies with dry air in the late postrainy season compared to postrainy season; 6- fifteen genotypes which did not flower in the postrainy season might be sensitive to high temperature and high atmospheric demands. These genotypes might be congenial only for the production of profuse vegetative growth rather than reproductive growth; 7- in all cases, GS₃ was shorter in the late postrainy. This might be due to prevailing high temperature, low relative humidity, longer day length, more hours of bright sunshine, clear skies and dry weather in the late postrainy season than in early postrainy season. All these unfavorable atmospheric conditions during late postrainy season might have led to reduction in grain size, quick grainfilling and low grain yield per panicle. Although the panicle size

did not decrease in size, grain number and grain weight were much lower. This indicated that the unfavorable weather caused abortion of grains during the late post-rainy season; 8- it may be assumed that high frequency irrigation was required in late postrainy season to cope with the high atmospheric demands prevailing in that season.

Table 5.11 Crop maturity of different sorghum genotypes in 2 seasons.

| Days to maturity | Postrainy season IS No. | Late postrainy season IS No. |
|------------------|---|---------------------------------|
| 80 | 633 | 8064 |
| 80-90 | 3951, 3460, 80644 7090, 3851, 3460, 301 9743, 1059, 13 | 310, 601, 3541, 127 |
| 90-100 | Bangkok tetraploid, 2210, 183, Bangkok, 183 4310, 601, 3541, 127, 7090, 474, 301, 474, 9743, 13, 658, 11150, 2445, 3818, 1059, 640, 2210 | |
| 100-110 | 3921, 4850, 640 658, 11150, 633, 4850, 2445 | |
| 110-120 | 3818 | |

Crop maturity:

The cultivars showed considerable variation for maturity periods within and between seasons. With a few exceptions, cultivars falling in group A, B and D matured much earlier in the rainy season than in the postrainy season. Cultivars showed shifts in their maturity status in the 2 seasons. A few cultivars (IS 474, IS 3460, IS 9743 and IS 4850) were fairly stable at the maturity stage in both seasons. During the postrainy season, many of the genotypes matured at 90-100 days, but a fairly large number did so between 80-90 days. Although many of the developmental stages were delayed in the late postrainy season, the maturity stage was reached early in many genotypes due to quick grainfilling. All the genotypes falling in group C (except IS 4850. IS 3921) were late in reaching physiological maturity. In other groups, with few exceptions, they matured earlier (Table 5.12).

Relationship among developmental stages (postrainy season 1976):

Some of the developmental stages showed significant correlations among themselves. Stem elongation phase was positively correlated with boot ($r=0.44$), half bloom ($r=0.43$) and soft dough ($r=0.46$). Flag leaf showed a significant negative relationship with panicle initiation ($r=-0.56$), flag leaf emergence ($r=-0.50$) and boot stage ($r=-0.46$). Days to anthesis showed high positive correla

tion with the total number of leaves ($r=0.86$) and stem elongation ($r=0.96$). Re *et al.* (1984) made a regression approach for prediction of sorghum phenology (GS₁, GS₂ & GS₃).

Yield components:

A common method of examining the potential of sorghum grain yield is to measure the total dry weight and dry grain yield and then compute the harvest index (HI):

$$\text{Harvest index (HI)} = (\text{Dry grain yield}) / (\text{Total dry weight})$$

$$\text{Dry grain yield} = (\text{HI}) \times (\text{Total dry weight})$$

A HI of 0.5 or more is an indication of high yields. Of all the components, the number of grains per branch is the most important. Panicle size, number of primary and secondary branches, grain weight, grain number per panicle, number of heads per unit area could be considered the yield components of sorghum and understanding their interrelationship is a key to improvement of sorghum. The degree of relationship among the yield components would determine the importance of a particular component. Weather conditions, cultural management and nutrient supply greatly influence yield components. Eastin *et al.* (1984) stated that development limitations seems to be critical in grain production under terms of high temperature stress. Aspects of yield components are discussed by different authors (Kambal and Webster, 1966; Stickler and Pauli, 1961; Blum, 1970a; Quinby, 1973; Beil and Atkins, 1967; Fischer and Wilson, 1975b). The effects of temperature on yield components are well documented (Tateno and Ojima, 1976; Chowdhury and Wardlaw, 1978). Grain number per head was significantly altered by temperature over the range of temperature (day and night) 21/16°C to 26/31°C and 30/25°C to 36/25°C; Chowdhury and Wardlaw, 1978; Downes 1972). Yields were markedly reduced at higher temperatures (30/25°C to 35/25°C) due to a reduction in grain weight (Tateno and Ojima, 1976). High temperatures during panicle development may reduce seed number per head and grain yield (Heinrich, 1981; Ogunlella, 1979).

Castleberry (1973) studied the effect of light energy available per plant on sorghum grain productivity by a series of thinnings. He observed that yield decreased until thinnings were done past floral differentiation (FD). Seed number per head increased sufficiently to maintain yield until FD, while the plant population was decreased by one-fourth (at about 2 weeks after PI). After this, population could not compensate in terms of seed number per unit of land area and increase in seed size could not compensate seed number loss (Eastin *et al.*, 1984). But Ogunlella (1979) demonstrated that weekly exposure to elevated night temperatures in the field, 5°C above ambient starting from PI, have reduced yield and seed number per head. Therefore the week after floral differentiation is the most sensitive to elevated night temperature. Dhopte (1984) demonstrated the deleterious effect of elevated night temperature on microsporogenesis and megasporogenesis, resulting in poor seed set. González-Hernández (1982) showed disruptive effects of temperature and water stress interactions in sorghum.

Eastin *et al.* (1984) stated that sorghum is relatively insensitive to heat stress during the vegetative stage, but stress have variable effects during panicle development, the most sensitive period being about 3 to 6 days after anthesis during microsporogenesis. Stress at post anthesis at 7 to 9 days cause restriction

in seed size and seed number.

Relationship among different panicle components: The number of primary branches showed less positive correlation with panicle length ($r=0.46$). The number of secondary branches was positively associated with the number of primary branches ($r=0.59$). Grain weight per panicle was highly associated with the number of secondary branches ($r=0.49$) and the number of grains per panicle ($r=0.55$; unpublished).

Relationship between growth stages with panicle components: During the summer, the different panicle components like the number of grains per panicle ($r=0.51$), weight of grains per panicle ($r=0.50$), 100 seed weight ($r=0.51$) and number of primary branches ($r=0.53$) were significantly associated with GS₃ duration ($P<0.01$). Stem elongation was found to be significantly correlated to GS₂ duration ($r=0.98$). Days to maturity was found to be a function of leaf number, flag leaf, GS₂ and stem elongation ($r^2=0.97$). GS₃ duration was a function of number of secondary branches, number of grains per panicle and grain weight per panicle ($r^2=0.36$). Grain number was found to be linearly correlated with the number of secondary branches ($Y = 17 + 0.13X$, $r^2=0.51$). Grain number was found to be a function of secondary branches, primary branches, GS₃ and grain weight ($r^2=0.58$). Grain size (100 seed weight) was a function of the rate of grain filling ($Y = 0.28 + 19.1X$, $r^2=0.89$). Grain weight showed relationship to secondary branches, grain number per panicle, primary branches and GS₃ ($r^2=0.39$). It was again found to be correlated to duration of grainfilling ($r=0.50$). Therefore, the rate of grainfill and duration of effective grainfill were both correlated with panicle productivity. Grain number and effective grainfilling period were not correlated with days to anthesis, which suggests that these characteristics were independent of maturity. Days to maturity was found to be a function of flag leaf stage, leaf number, GS₂ and stem elongation ($r^2=0.93$; unpublished).

GENERAL COMMENTS

The transformation of the vegetative apex to the reproductive apex is the most important phenomenon in the life cycle of cereals. Temperature and photoperiod are the 2 important factors controlling this internal autonomous progression, when crops are grown under optimum soil moisture and nutrient conditions. This change in the formation of a completely dissimilar structure is a hormone controlled phenomenon and guided by photothermal interactions prevailing in the growing season of the crop. This phase represents the cessation by 1 group of genes and the initiation of activity by another (Milthorpe and Moorby, 1976).

Most sorghum genotypes seem to be short day plants and on the basis of their response to day length, they can be grouped into photoperiod-insensitive, obligate photoperiod-sensitive, and facultative photoperiod-sensitive. Photothermal interactions play an important role in controlling flowering of these 3 groups. For this reason, temperate sorghum behave differently when grown under tropical environments. To break this tropical-temperate barrier, a suitable crossing program should be adopted in order to develop cultivars for broad adaptations.

This approach could give greater impetus to the sorghum crop improvement program. In the tropics, photosensitive sorghum grows very tall in the rainy season and often produces a large number of leaves and a few productive heads. To avoid this, photoinensitive sorghum are generally preferred.

As yield is the primary goal of the breeder, optimum growing conditions are essential for the expression of a crop's full genetic potential. Significant yield improvement has been achieved in different crops, but only under high input situations. In these favorable environments, the potential yield is determined during the early stage of inflorescence development and the panicle meristem is capable of producing the optimum number of florets to their full genetic potential. This is simultaneously substantiated by the optimum photosynthetic efficiency of the leaf attained during the vegetative stage. Nevertheless, it is very difficult to provide congenial growing conditions in most semiarid regions of the world. Often, crops are prevented from expressing their full genetic potential. The productivity of the panicle is drastically reduced due to several unfavorable environmental conditions like lack of water, low nutrients availability, salinity, high temperature, etc. Therefore, major research efforts need to be directed to screen genotypes which could express their optimal genetic potential under unfavorable field conditions.

In substantiation of this, under water stress situations, the growth and development of panicles is affected to an extent by the intensity of water stress at different stages of development. Under severe water stress when cell division is of paramount importance, spikelet differentiation tends to stop but under moderate stress situation, differentiation may be delayed not suspended. Under severe water stress, failure of stem and internode elongation leads to poor or no exertion of the panicle. There could be a drastic reduction in floret number, thereby impairing the final productivity of the panicle and crop. Water stress affects the germination of pollen and growth of pollen tubes on stigmas. During grain development, water stress affects the sustained growth rate of grain. This ultimately leads to poor seed setting and filling of the grain and drastic reduction in seed size. It also has a direct effect on seedling establishment (Chapter-3). Therefore, drought avoidance trait with its capability to complete panicle development and anthesis within a short span of time is an adaptive mechanism. Lines could be selected from the group which express better panicle growth. These could be incorporated in the crop improvement program.

A full understanding of the reproductive apex and its interaction with environmental factors could provide some clues for the selection of cultivars and genetic improvement of the crop.

The maximum yield is determined by the potential of a crop variety and its adaptation to a particular environment. At present, breeders look for a dwarf plant with a compact panicle. However, in a tropical environment, compact panicle provides a favorable environment for the infestation and disease, like grain moths and earhead bugs. A lax panicle provides less opportunity for the development of the biotic stress. The focus of research should be to increase grain number per panicle and grain weight rather than evolving a dwarf plant prone to infestation. An ideotype in sorghum needs to be formulated for better adaptation in the environment.



ROOT DEVELOPMENT AND GROWTH

INTRODUCTION

Physiologists are more interested in the above ground portions of plants which play the central role during photosynthesis in plant metabolism. Very little attention is given to studies on the growth and function of root systems. This is partly due to the unavailability of an efficient technique for the extraction of roots from the field. Considerable variation in the expression of the size of roots occurs under different environmental conditions (Russell, 1977). Due to the difficulties involved, most research endeavors concerning root studies are conducted in greenhouses, growth chambers, rhizotrons and a few are based on observations in the field (Kaigama *et al.*, 1977). It is not possible to make a complete analysis of the growth and function of either roots or shoots unless the interrelationship between them and their behaviour in edaphic systems is taken into account (Russell, 1977).

Different techniques have been adopted to study root development in different crops. In sorghum, soil cores are taken from the field with the help of tubes to study depthwise distribution of roots and to correlate the relationship between root and the above-ground portion of the plant at different morphological development stages. Root morphogenesis and early growth have been done mostly in hydroponic culture (Blum *et al.*, 1977 a,b), sand culture (Nour and Weibel, 1978), and glass house pot culture (Hackett, 1973). Böhm (1977) described and compared different methods of root observation, both destructive and nondestructive in a natural environment in mini-rhizotrons. The use of clear tubes buried in the soil, or mini-rhizotron, has been described by different authors (Waddington, 1971; Böhm, 1977). Böhm (1977) stated that mini-rhizotrons required less time for data collections compared to other methods. Other methods have been adopted for the study of root system in different crops (Newmann, 1966; Marsh, 1971; Tennant, 1975; Voorhees, 1976; Sanders and Brown, 1978; Foale and Upchurch, 1982).

An endoscope introduced into a transparent tube and placed into the soil before sowing has been used for direct observation of root distribution and intensity of root colonization (Martens and Clauzel, 1982).

The root system of sorghum plants grown in soil measured at 9, 14 and 17 days from sowing indicate the low average diameter of the root number and the very high extension rate. Sorghum appeared to maintain a stable relationship between the overall number, length, surface area and volume of its root (Hackett, 1973). In sorghum, both under irrigated and non-irrigated conditions a rapid penetration